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Dekker, Nynke; Ha, Seungkyu; Janissen, Richard; Ussembayev, Yera; van Oene, Maarten; Solano Hermosilla, Belen

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Tunable top-down fabrication and functional surface coating of single-crystal titanium dioxide nanostructures and nanoparticles†

Seungkyu Ha, Richard Janissen, Yera Ye. Ussembayev, Maarten M. van Oene, Belen Solano and Nynke H. Dekker*

Titanium dioxide (TiO2) is a key component of diverse optical and electronic applications that exploit its exceptional material properties. In particular, the use of TiO2 in its single-crystalline phase can offer substantial advantages over its amorphous and polycrystalline phases for existing and yet-to-be-developed applications. However, the implementation of single-crystal TiO2 has been hampered by challenges in its fabrication and subsequent surface functionalization. Here, we introduce a novel top-down approach that allows for batch fabrication of uniform high-aspect-ratio single-crystal TiO2 nanostructures with targeted sidewall profiles. We complement our fabrication approach with a functionalization strategy that achieves dense, uniform, and area-selective coating with a variety of biomolecules. This allows us to fabricate single-crystal rutile TiO2 nanocylinders tethered with individual DNA molecules for use as force- and torque-transducers in an optical torque wrench. These developments provide the means for increased exploitation of the superior material properties of single-crystal TiO2 at the nanoscale.

Introduction

Micro- and nanostructures based on titanium dioxide (TiO2) have been utilized in numerous applications that exploit the unique properties of this material (Fig. S1†). For example, the high photocatalytic activity of TiO2 is used for water splitting1 and air purification.2 The electrical properties of TiO2, which include a tunable resistance, a high dielectric constant, carrier transport abilities, and a wide band gap, render it a preferred material for sensors,3 memory devices,4 transistors,5 solar cells,6 and batteries.7,8 Furthermore, the optical properties of TiO2, e.g. high refractive index and optical nonlinearity, make it find use in optical waveguides,9 photonic crystals,10 and optical trapping.11

In particular, the use of TiO2 in its single-crystalline phase can offer substantial advantages over its amorphous and polycrystalline phases for existing and new applications. For instance, single-crystal TiO2 allows improved control of surface properties, e.g. photochemical reactivity12 and surface wettability,13 by making use of the different crystallographic orientations. The higher carrier mobility of single-crystal TiO2 renders it a promising material for transistors and (bio)sensors14 and may enhance the efficiency of dye-sensitized solar cells.15 Furthermore, optical waveguides and photonic crystals composed of single-crystal TiO2 are expected, due to their highly ordered atomic structures, to exhibit improved light transmission and nonlinear responses.18,19 Lastly, the single-crystal rutile polymorph of TiO2 has an exceptionally large optical birefringence, which makes it an excellent candidate for incorporation into torque transducers for e.g. single-molecule spectroscopy.20

To facilitate the application of TiO2 in its single-crystalline phase, major challenges in both fabrication and surface functionalization must be addressed. First, it is important to control structural parameters such as feature size, shape, and pitch that determine the resulting physical responses of fabricated TiO2 nanostructures. Such physical responses impact e.g. gas sensing performance,3 light scattering efficiency,6 light propagation behavior,9 and optical trapping capabilities.11

Department of Bionanoscience, Kavli Institute of Nanoscience, Faculty of Applied Sciences, Delft University of Technology, Lorentzweg 1, 2628 CJ Delft, The Netherlands. E-mail: n.h.dekker@tudelft.nl
†Electronic supplementary information (ESI) available: Experimental details (ESI Methods) of the optic axis orientation of TiO2 nanocylinders, Cr etch mask fabrication, surface functionalization and its evaluation using fluorescence microscopy, preparation of DNA constructs, assembly of flow cells, bioconjugation of TiO2 nanocylinders, OTW instrumentation and measurements; TiO2 dry etching optimization and the etching parameters employed (Tables S1 and S2); dimensional analysis of TiO2 nanocylinders (Table S3); diverse applications of TiO2 at the nanoscale (Fig. S1); selection of etch mask material (Fig. S2); control of sidewall profiles in TiO2 etching (Fig. S3); size distributions of TiO2 nanocylinders (Fig. S4); quantitative comparisons of different surface linker molecules (Fig. S5); DLS measurements on TiO2 nanocylinders (Fig. S6); optical trap calibration (Fig. S7); and supplementary references. See DOI: 10.1039/c6nr00898d
Mainstream bottom-up approaches for TiO₂ nanofabrication, e.g. the hydrothermal growth method and electrochemical anodization, although capable of high throughput, lack sufficient flexibility in controlling dimensions and geometries. A number of bottom-up methods are capable of fabricating single-crystal TiO₂ nanostructures, but these lack control of the crystallographic orientations and are limited to a certain crystalline polymorph, e.g. anatase or rutile. Hybrid fabrication methods, such as the atomic layer deposition array defined by etch-back technique, can achieve a high aspect-ratio but are limited to the fabrication of amorphous or polycrystalline TiO₂. These limitations can be overcome by employing top-down nanofabrication methods based on plasma etching, which can target a single-crystal TiO₂ substrate with any desired crystalline polymorph or crystallographic orientation. However, as TiO₂ is substantially more difficult to etch than comparable materials e.g. silicon (Si), silicon dioxide (SiO₂), and silicon nitride (Si₃N₄), such etching processes are currently limited to a low aspect-ratio and a largely positive sidewall angle.

Second, it is key to achieve highly efficient surface functionalization and bioconjugation of TiO₂ for applications such as (bio)sensors, force transducers, and torque transducers. However, surface functionalization of TiO₂ using common alkoxysilane surface linkers, is known to be less efficient in terms of linker density and uniformity than other oxide materials e.g. SiO₂ and aluminum oxide (Al₂O₃). Furthermore, the majority of the described bioconjugation methods for TiO₂ surfaces require a long incubation time and additional crosslinker molecules that could decrease the overall conjugation efficiency and harm the biomolecule functionality. Peculiarly for single-crystal TiO₂ nanostructures, the lack of extensive quantitative and qualitative comparisons of different surface functionalization strategies hinders the selection of an optimal procedure.

Here, we present a successful top-down fabrication and surface functionalization of single-crystal TiO₂ nanostructures and nanoparticles. We focus our efforts on the rutile poly-crystal TiO₂ (100), in order to harness its exceptionally large optical birefringence and precisely oriented optic axis for effective torque transfer (ESI Methods†) in an optical torque wrench (OTW). Using high aspect-ratio nanocylinders suitable for optical trapping as our target structures, we demonstrate how these can be mass-produced with well-defined sizes and etch profiles that display excellent overall structural uniformity. We achieve a dense, uniform surface coating of TiO₂ nanocylinders using different linker molecules, and we demonstrate subsequent covalent conjugation of biological and organic molecules such as DNA, streptavidin, biotin, and biocompatible poly(ethylene glycol) (PEG). We successfully exploit these developments to fabricate and functionalize TiO₂ nanocylinders with single DNA molecules, and demonstrate full rotational control of individual nanocylinders in an OTW. This provides a particularly stringent test, as proper functioning of the OTW relies on both the correctly oriented optic axis and tightly controlled dimension of the single-crystal TiO₂ nanocylinder, and the quality and reliability of the surface coating. Rutile being the polymorph of TiO₂ that is most difficult to etch, our approach provides the means for increased exploitation of the superior material properties of single-crystal TiO₂ in any desired polymorph for diverse applications.

Results and discussion

Selecting an etch mask for the plasma etching of single-crystal TiO₂

One of the major issues in top-down nanofabrication of single-crystal TiO₂ is its high etch resistance. Plasma generated by fluorine-based gases is capable of etching TiO₂ only under more aggressive etching conditions – in terms of plasma ion density and energy – than the ones employed for the dry etching of comparable materials e.g. Si, SiO₂, and Si₃N₄. The increased difficulty in etching TiO₂ compared to e.g. SiO₂ stems from differences in the volatility of their etch byproducts: whereas SiF₄ is volatile, TiF₄ is not. Also, masks can suffer from excessive edge erosion during etching, which in turn reduces both feature sizes and the ability to perform anisotropic etching.

To overcome these challenges, we have searched for a mask material for the dry etching of single-crystal TiO₂ that achieves sufficiently high etch selectivity to permit the fabrication of nanostructures in a wide range of dimensions and sidewall profiles. We have compared different mask materials (Fig. S2†), such as chromium (Cr), tungsten (W), e-beam resist, and amorphous Si. We observe that in particular Cr provides a significantly higher etch selectivity in fluorine-based etching of rutile TiO₂ (up to ~28:1 under specific etching conditions; an overview of etch selectivity values for different etching conditions can be found in Fig. S3†) than the other tested materials. We therefore select Cr as an appropriate etch mask material. To fabricate the Cr mask, we create circular apertures in a poly(methyl methacrylate) (PMMA) layer using e-beam lithography (Fig. 1, steps 1–3). The aperture diameters can be tuned within 100–600 nm by means of defocusing e-beam. Subsequent Cr deposition and the following lift-off define the mask (Fig. 1, steps 4 and 5).

Trifluoromethane (CHF₃)-based plasma etching of single-crystal TiO₂ nanocylinders

With the objective to maximize the etch rate with a sufficiently high etch selectivity (>10:1), in order to achieve etch depths up to 1–2 μm at the practical upper limit of Cr mask thickness, ~150 nm; ESI Methods†) and to produce single-crystal TiO₂...
nanocylinders with vertical sidewall angles, we have optimized parameters for the fluorine-based plasma etching process (Fig. 1, step 6). We start with a reactive ion etching (RIE) system in which trifluoromethane (CHF$_3$) is the major etchant, and oxygen (O$_2$) and argon (Ar) are additive gases. The major etchant gas CHF$_3$ supplies both the reactive etching species and the fluorocarbon ($C_{x}F_{y}$) surface passivation layer-forming species. To optimize the process, we systematically vary the radio frequency (RF) power, chamber pressure, and ratios of the etch gases (CHF$_3$ : O$_2$ : Ar) (Table S1†). Our optimized etching conditions (Table S2†) result in a high single-crystal TiO$_2$ etch rate of ~40 nm min$^{-1}$ at a sufficient etch selectivity of ~14 : 1 over the Cr mask, which enables the formation of TiO$_2$ nanocylinders with vertical sidewalls (~90°; Fig. 2a). We have successfully implemented this protocol on a second, nominally identical RIE system (Fig. 2b), requiring only minor tuning in the O$_2$ flow rate (4–8 sccm compared to ~0.5 sccm), most likely due to differences in instrument calibration.

Notably, we observe that tuning a single process parameter, namely the O$_2$ flow rate, allows us to drastically alter sidewall etch profiles and even cross-sectional shapes (Fig. S3†). For example, at low O$_2$ flow rates (0–1 sccm), we can modulate the sidewall angles from positive to negative, thereby including vertical sidewalls (Fig. 2a and b). At significantly higher O$_2$ flow rates (5–10 sccm), we can obtain hourglass-shaped etch profiles (Fig. 2c).

**Sulfur hexafluoride (SF$_6$)-based plasma etching of single-crystal TiO$_2$ nanocylinders**

While maintaining identical objectives as above (high etch selectivity and vertical sidewall angles), we have also optimized the plasma etching of TiO$_2$ using another major etchant,
profiles. The higher TiO$_2$ etch rates that we obtain result from different conditions in plasma etching. In each SEM image, the top surface of the nanocylinder is marked by a yellow line (remnants of the Cr mask prior to its removal are visible in (b, c and f)). Scale bars denote 500 nm. In the following, the height ($H$), the averaged diameter ($D$), and the resulting aspect-ratio ($AR = H/D$) of each nanocylinder are shown. (a) A nanocylinder with a vertical sidewall ($89^\circ$) that results from an optimized CHF$_3$:O$_2$ dry etching in an ICP-RIE system ($H$: 430 nm, $D$: 110 nm, AR: 3.9). (b) A nanocylinder with a vertical sidewall ($89^\circ$) etched from the optimized CHF$_3$:O$_2$ dry etching in a second, nominally identical RIE system ($H$: 605 nm, $D$: 205 nm, AR: 3.0). (c) An hourglass-shaped nanocylinder that results from CHF$_3$:O$_2$:Ar dry etching in an RIE system ($H$: 1020 nm, $D$: 200 nm, AR: 5.1). (d) A cone-shaped nanocylinder that results from SF$_6$:CH$_4$:Ar dry etching in an ICP-RIE system ($H$: 1385 nm, $D$: 490 nm, AR: 2.8). (e) A cone-shaped nanocylinder that results from SF$_6$:He dry etching in an ICP-RIE system ($H$: 2160 nm, $D$: 390 nm, AR: 5.5). (f) A nearly vertical ($85^\circ$) nanocylinder that results from an optimized SF$_6$:He dry etching in an ICP-RIE system ($H$: 470 nm, $D$: 210 nm, AR: 2.2).

Quantification of single-crystal TiO$_2$ nanocylinder uniformity

To quantify both the local and global structural uniformity of fabricated single-crystal TiO$_2$ nanocylinders over an entire substrate, we have analyzed SEM images of high aspect-ratio (3.6) nanocylinders fabricated using the CHF$_3$ plasma-based protocol. From the analysis of top-view SEM images (Fig. 3b), we observe that the top (Fig. 3d) and bottom diameters (Fig. 3e) exhibit excellent local uniformity, as defined as the uniformity at each measurement position (averaged relative standard deviation (RSD) of 4.4% (top) and 2.3% (bottom)). The global uniformity across the substrate is shown by SEM images (Fig. 3c), we also extract the nanocylinder heights (Fig. 3f), which similarly show very good uniformity both locally (averaged RSD of 0.6%) and globally (RSD of 1.4%). We attribute the nearly monotonic increase in heights outward from the substrate center to etch rate nonuniformity and the fact that etching erodes the Cr mask in a non-uniform fashion depending on the roughness of the mask surface, as backed up by an analysis of the roughness of the top and bottom nanocylinder surfaces. From tilted-view SEM images (Fig. 3f), we also extract the nanocylinder heights (Fig. 3f), which similarly show very good uniformity both locally (averaged RSD of 0.6%) and globally (RSD of 1.4%). We attribute the nearly monotonic increase in heights outward from the substrate center to etch rate nonuniformity. The analysis of a batch of low aspect-ratio (1.6) nanocylinders reveals identical trends (Fig. S4 and Table S3;† etching conditions in Table S2†), thereby proving that our protocol can be used to generate both low and high aspect-ratio nanocylinder shapes with high uniformity.
Surface functionalization and bioconjugation of single-crystal TiO$_2$

We now demonstrate highly efficient and versatile surface functionalization and bioconjugation of single-crystal rutile TiO$_2$ (Fig. 4a). We perform the functionalization experiments on single-crystal TiO$_2$ nanostructures fabricated using our developed SF$_6$-based plasma etching process presented earlier. Compared with CHF$_3$-based etching, this process results in a lower degree of surface contamination by C$_x$F$_y$, and hence a higher surface linker coverage.

To quantitatively characterize the surface coating efficiency of different covalently bound surface linkers, we first test functionalization on single-crystal rutile TiO$_2$ substrates with large-scale square micro-patterns ($25 \times 25$ $\mu$m$^2$) embedded in a thick (1–2 $\mu$m) PMMA layer (Fig. 4b). Similarly to other oxide materials, the surface of TiO$_2$ is populated by hydroxyl groups that allow it to be functionalized with different types of linker molecules, such as silanes and other organic molecules with a covalent hydroxyl-binding capacity.$^{33,41}$ Here, we have tested four different surface linker molecules: an aminoalcohol (ethanolamine hydrochloride; ETA), an epoxysilane ((3-glycidoxypropyl)dimethylethoxysilane; GPDMES), a widely used alkoxy silane ((3-aminopropyl)dimethylethoxysilane; APDMES), and a cyclic azasilane (N-$n$-butyl-aza-2,2-dimethoxysilacyclo-pentane; BADMSCP). Additionally, since PEG coatings are widely used for surface passivation$^{42}$ and as biocompatibility layers,$^{43}$ we have also tested the covalent attachment of heterobifunctional PEG to ETA-coated surfaces (ESI Methods, Fig. S5†). To compare the coating efficiency of these different linkers, we covalently bind fluorophores (ATTO 647N; $\lambda_{\text{excitation}} = 640$ nm, $\lambda_{\text{emission}} = 669$ nm) to the functionalized micro-patterns and measure the fluorescence intensity using quantitative fluorescence microscopy (ESI Methods†). The fluorescence measurements report successful surface functionalization for all the surface linkers used (Fig. S5†).

Regarding subsequent bioconjugation to TiO$_2$ surfaces, several considerations led us to select epoxysilane (GPDMES) as the most appropriate surface linker. To start, epoxysilanes are widely used for their demonstrated reliability in surface functionalization$^{44}$ and biomolecule attachment.$^{45}$ Furthermore, GPDMES-coated TiO$_2$ surfaces demonstrate high surface linker coating density and homogeneity (Fig. 4b). Indeed, while GPDMES coatings provide a slightly lower surface linker
coated TiO2 substrate. The corresponding zoom-in amino-labeled (right panel) of a square shows successful covalent binding of the structures for the qualitative characterization of the e lized surface. Scale bars denote 20 µm. (c) Functionalized nano-cylinders using the same method as above. Scale bars denote 2 µm.

Fig. 4 Characterization of the efficiency of surface functionalization of single-crystal TiO2 structures using fluorescence microscopy. (a) Illustration of the surface coating steps. First, single-crystal TiO2 surfaces are O2 plasma-treated to generate hydroxyl groups (—OH). Then epoxy-silanes (GPDMES) are covalently bound to the surface hydroxyl groups. Consequently, either fluorophores (red) or biomolecules (e.g., streptavidin (green)) are bound covalently to the reactive epoxy functional group. (b) Functionalized micro-patterns for the quantitative characterization of the efficiency of surface functionalization. The brightfield image (left panel) shows squares (25 x 25 µm²) patterned on the PMMA-coated TiO2 substrate. The corresponding zoom-in fluorescence image (right panel) of a square shows successful covalent binding of the amino-labeled fluorophores (ATTO 647N) to the epoxysilane functionalized surface. Scale bars denote 20 µm. (c) Functionalized nano-structures for the qualitative characterization of the efficiency of surface functionalization. Brightfield (left panel) and the corresponding fluorescence (right panel) images of top-area functionalized TiO2 nanocylinders using the same method as above. Scale bars denote 2 µm.

density than ETA or PEGylated ETA coatings, their coating efficiency is significantly higher than the one obtained using BADMSCP and comparable to that of commonly used APDMES (Fig. S5†).

More importantly, the greatest advantages of using GPDMES compared to other surfactant linkers are: first, the ability of the epoxy group to react directly with the primary amino groups of biomolecules that permits bioconjugation in the absence of additional crosslinkers, avoiding conditions that could harm the structural and functional integrity of the target biomolecules; and second, the rapid surface functionalization process (e.g., ~15 min compared to ~12 h of all the other linkers; ESI Methods†). With this procedure in hand, epoxysilane-functionalized TiO2 surfaces can be readily used for the covalent conjugation of a majority of biomolecules. We apply this bioconjugation process to the single-crystal TiO2 nanocylinder arrays on the substrate (Fig. 1, steps 7–11) prior to their mechanical cleaving (Fig. 1, step 12) to avoid additional, subsequent purification procedures. At this stage, functionalization can be applied to specific areas of the TiO2 nanocylinders through partial PMMA coating (Fig. 1, steps 8–10). Using this approach, we are able to functionalize and bioconjugate only the top parts of the TiO2 nanocylinders (Fig. 1, step 11). Fluorescence microscopy has also been used to qualitatively demonstrate the successful functionalization of individual TiO2 nanocylinders (Fig. 4c).

Degree of monodispersity in coated single-crystal TiO2 nanoparticles

Our ability to functionalize single-crystal TiO2 nanoparticles with high surface linker density and homogeneity can also be beneficial in different contexts. For example, following the mechanical cleaving that liberates the nanocylinders from the TiO2 substrate, it is necessary to prevent their aggregation in aqueous environments. Generally speaking, the aggregation of nanoparticles depends on both the surface composition of the nanoparticles and the type of solution in which they are immersed.46 We have tested different surface coatings and buffer solutions that effectively prohibit single-crystal TiO2 nanoparticle aggregation and have evaluated these using dynamic light scattering (DLS) measurements (Fig. S6†). We found that non-coated TiO2 nanocylinders aggregate substantially over time in deionized (DI) water, in contrast to what occurs in physiological phosphate buffered saline (PBS) solution at a similar pH (7.4). We also found that the addition of bovine serum albumin (BSA, 2% (m/v)), which is widely used as a surface passivation agent,47 deteriorates substantially the monodispersity of TiO2 nanocylinders in both DI water and PBS solution. When we apply a PEG coating – widely implemented for passivating nanoparticle surfaces to decrease aggregation and for enhancing their biocompatibility43 – to TiO2 nanocylinders, we observe no aggregation in either DI water or PBS solution. Such PEG-coated TiO2 nanocylinders refrain from aggregation even when they are conjugated to biomolecules such as biotin or DNA. Likewise, the GPDMES-coated TiO2 nanocylinders with bioconjugated streptavidin show no aggregation under physiological conditions (PBS solution) and therefore have been chosen for the single-molecule OTW experiments.

Characterization of individual DNA molecules using single-crystal TiO2 nanocylinders

To demonstrate the potential of our developed fabrication and surface coating processes for diverse applications, we tether single-crystal TiO2 nanocylinders to individual linear, double-stranded DNA molecules and perform stretching and twisting experiments in an OTW. For these measurements, we use
GPDMES-functionalized, streptavidin-coated single-crystal TiO₂ nanocylinders fabricated in SF₆ plasma (Methods), and test their response to force and torque applied on torsionally constrained 20.6 kbp DNA (contour length of \( \sim 7 \mu m \); ESI Methods†). A biotinylated handle on one extremity of the DNA is designed to bind to the streptavidin-coated nanocylinders, whereas the other extremity of the DNA includes digoxigenin to allow it to bind to the digoxigenin antibodies that are covalently bound to a heterobifunctional PEG-coated flow cell channel (Fig. 5a, ESI Methods†).

Prior to performing measurements on these DNA-tethered single-crystal TiO₂ nanocylinders in an OTW (ESI Methods†), we calibrate the optical trap by using individual non-tethered nanocylinders to measure its trap stiffness (Fig. S7†). We can then apply a force to a tethered nanocylinder and stretch the DNA (Fig. 5b). The obtained force-extension data are fitted to the worm-like chain (WLC) model⁴⁸ (red line in Fig. 5b) and yield a persistence length of 45 ± 1 nm and a contour length of 6.8 ± 0.2 \( \mu m \) (means and standard deviations for \( n = 5 \) different DNA tethers), in excellent agreement with previous reports.³₁,⁴⁹–⁵² We note that the OTW provides an excellent platform to evaluate the quality of our top-down fabrication of single-crystal TiO₂ nanostructures, as only nanocylinders with precisely controlled dimensions and uniformity are appropriate for stable and reproducible optical trapping. Furthermore, the use of the OTW to stretch and twist DNA molecules provides a stringent test of the applied surface functionalization, as these processes (notably twisting) are only possible provided that the surface functionalization and bioconjugation processes are sufficiently efficient to allow for the formation of multiple stable bonds at either DNA extremity.⁵²

Conclusions

We have developed and optimized processes for the fabrication, surface functionalization, and bioconjugation of single-crystal TiO₂ nanostructures and nanoparticles. Using two top-down etching approaches, we demonstrate the fabrication of highly uniform nanostructures and nanoparticles with controllable shapes and dimensions. The CHF₃-based etching process provides a wider range of etch geometries, while the SF₆-based etching process results in improved surface quality allowing homogeneous and dense coating. We are able to etch single-crystal TiO₂ into various nanoscale shapes, including cylinders, cones, and hourglass-shaped structures. The fabricated
nanostructures have diameters in the range of 100–600 nm, with heights up to 1–2 μm under etching conditions that yield high etch rate and selectivity. Using SF$_6$-etched single-crystal TiO$_2$ nanocylinders, we show how they can be functionalized with high efficiency and be conjugated covalently to a majority of biomolecules. We demonstrate the use of these coated TiO$_2$ nanocylinders by stretching and twisting individually tethered DNA molecules, an application that exploits the tight dimensional control and high quality surface functionalization of nanocylinders etched from a rutile (100) single-crystal TiO$_2$ substrate. Our developed fabrication and surface functionalization methods will likely find utilization in diverse optical and electronic applications of single-crystal TiO$_2$, including applications in life science-oriented fields through e.g. the development of biosensors and single-molecule techniques.

**Methods**

**Fabrication and bioconjugation of single-crystal rutile TiO$_2$ nanocylinders for OTW experiments**

We use single-side polished single-crystal rutile TiO$_2$ (100) substrates (Latech, Singapore), 10 × 10 mm$^2$ in size and 0.5 mm in thickness, to produce nanocylinders. We clean the substrates in fuming nitric acid (99.5%, Sigma-Aldrich, The Netherlands) using ultrasonication for 10 min followed by thorough rinsing in DI water and drying under a nitrogen (N$_2$) stream (Fig. 1, step 1). We then subject the substrates to an O$_2$ plasma (TePla300, PVA TePla, Germany) at 500 rpm for 5 s and then 3000 rpm for 55 s. We bake the resist-coated substrates on a hotplate for 30 min at 175 °C (Fig. 1, step 2). The substrates do not require an additional conductive layer to prevent charge accumulation during e-beam patterning since it has sufficiently high electrical conductivity.

We then use an e-beam patterning system (EBPG 5000+, Leica, UK; the acceleration voltage is set to 100 kV and the aperture diameter to 400 μm) to pattern the PMMA layer by a defocused e-beam with circular cross-section and tunable diameter. The patterning of each circular shape is performed by a single-pixel e-beam exposure, defining the shapes of the Cr mask for subsequent nanocylinder etching. Using optimized e-beam conditions (a current of 85 nA, a defocusing of 100–200 nm, and a dose of 2000 μC cm$^{-2}$), we pattern circles with 600 nm pitch on a circular patterning area with radius ~4 mm (~140 million masks in total) in ~30 min. We develop the patterned substrate using a solution of methyl isobutyl ketone (MIBK, Sigma-Aldrich, The Netherlands) and isopropyl alcohol (IPA, Sigma-Aldrich, The Netherlands) (1 : 3 MIBK : IPA) for 60 s with constant hand agitation at room temperature. We then rinse the substrate in IPA for 30 s and dry it under a N$_2$ stream (Fig. 1, step 3).

We deposit the Cr mask layer using either an e-beam evaporator (Temescal FC-2000, Ferrotec, Germany) or a thermal evaporator (L560, Leybold Heraeus, Germany) loaded with Cr pieces (purity of 99.95%, Kurt J. Lesker, Germany), using the patterned PMMA layer as evaporation windows. We use slow Cr deposition rates ($\leq$1.0 Å s$^{-1}$) to obtain high quality Cr layers (Fig. 1, step 4). We finalize Cr mask formation in a lift-off process by soaking the substrate into a preheated (80 °C) resist stripper (PRS-3000, JT Baker, The Netherlands) for 20 min with constant agitation using a magnetic spinner. Then we thoroughly rinse the substrate in DI water and dry it under a N$_2$ stream (Fig. 1, step 5).

We etch TiO$_2$ nanocylinders using the optimized conditions (Table S2†) for vertical sidewall angles (Fig. 1, step 6). We etch nanocylinders with either an RIE system (Flour Z401S, Leybold Heraeus, Germany; Fig. 2a and b) or an ICP-RIE system (Plasmalab system 100, Oxford Instr., UK; Fig. 2f). For dimensional analysis of the etched nanocylinders, we employ SEM systems (Hitachi S4800 FESEM, FEI XL30S FEG SEM, or FEI Nova NanoSEM). We remove the used Cr mask using a wet etchant (Chromium Etchant N° 1, MicroChemicals, Germany) and then perform surface cleaning using fuming nitric acid and O$_2$ plasma (TePla300) for the subsequent surface functionalization (Fig. 1, step 7).

Optionally, if area-selective functionalization on top of the nanocylinders is required, we cover their sidewalls with a temporary protection layer. We cover the substrate with a PMMA 950k resist at a chosen concentration to provide a spin-coated thickness larger than the heights of the nanocylinders (Fig. 1, step 8). Consequently, we etch the PMMA layer to render the tops of the nanocylinders accessible, using either O$_2$ plasma (TePla300) or a MIBK solution (Fig. 1, step 9). Note that we do not bake the resist in order to allow for its straightforward removal afterwards.

To attach biomolecules to TiO$_2$ nanocylinders, we first activate the nanocylinder substrates by O$_2$ plasma treatment (Plasma-PREEN I, Plasmatic Systems Inc., USA). Then we bind GDPMES to the substrate for 15 min at 75 °C using non-diluted GDPMES solution, and wash the substrate in chloroform (CHROMASOLV plus, for HPLC, ≥99.9%, Sigma-Aldrich, The Netherlands) three times for 15 s each, followed by drying under a N$_2$ stream (Fig. 1, step 10). (If a temporary PMMA layer is used for area-selective functionalization, we first wash the substrate in ethanol three times for 15 s each directly after the incubation with GDPMES, followed by drying under a N$_2$ stream. We then remove the temporary PMMA layer in chloroform for 15 min at room temperature, followed by an additional washing in fresh chloroform three times for 15 s each and drying under a N$_2$ stream.)

We then bind streptavidin molecules to the epoxy group of GDPMES by incubating the nanocylinders with 2 μM streptavidin (Sigma-Aldrich, The Netherlands) in PBS solution (pH 7.4, Sigma-Aldrich, The Netherlands) for 1 h at room temperature. After washing the substrate three times for 15 s each with PBS/TWEEN$^\text{®}$ solution (pH 7.4, PBS containing 0.1% v/v TWEEN$^\text{®}$ 20 (Sigma-Aldrich, The Netherlands)), we wash the substrate with PBS solution (pH 7.4) another three times for 15 s each (Fig. 1, step 11).

We cleave TiO$_2$ nanocylinders mechanically from the substrate using a diamond microtome blade (DT315D50, C.L. Nanosc.
Sturkey, USA) inside a PBS/Triton™ droplet (pH 7.4, PBS containing 0.05% (v/v) Triton™ X-100 (Sigma-Aldrich, The Netherlands)), followed by suction of the droplet using a micropipette (Fig. 1, step 12). These cleaved, streptavidin-coated TiO₂ nanocylinders can be further coupled with biomolecules of interest, e.g. DNA with biotinylated handles, by incubating them in a flow cell channel for OTW experiments (ESI Methods†).

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