

## Influence of neoadjuvant chemotherapy on diffuse reflectance spectra of tissue in breast surgery specimens

De Boer, Lisanne L.; Kho, Esther; Józwiak, Katarzyna; Van De Vijver, Koen K.; Vrancken Peeters, Marie Jeanne T.F.D.; Van Duijnhoven, Frederieke; Hendriks, Benno H.W.; Sterenborg, Henricus J.C.M.; Ruers, Theo J.M.

**DOI**

[10.1117/1.JBO.24.11.115004](https://doi.org/10.1117/1.JBO.24.11.115004)

**Publication date**

2019

**Document Version**

Final published version

**Published in**

Journal of Biomedical Optics

**Citation (APA)**

De Boer, L. L., Kho, E., Józwiak, K., Van De Vijver, K. K., Vrancken Peeters, M. J. T. F. D., Van Duijnhoven, F., Hendriks, B. H. W., Sterenborg, H. J. C. M., & Ruers, T. J. M. (2019). Influence of neoadjuvant chemotherapy on diffuse reflectance spectra of tissue in breast surgery specimens. *Journal of Biomedical Optics*, 24(11), Article 115004. <https://doi.org/10.1117/1.JBO.24.11.115004>

**Important note**

To cite this publication, please use the final published version (if applicable).  
Please check the document version above.

**Copyright**

Other than for strictly personal use, it is not permitted to download, forward or distribute the text or part of it, without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license such as Creative Commons.

**Takedown policy**

Please contact us and provide details if you believe this document breaches copyrights.  
We will remove access to the work immediately and investigate your claim.

## **Influence of neoadjuvant chemotherapy on diffuse reflectance spectra of tissue in breast surgery specimens**

Lisanne L. de Boer  
Esther Kho  
Katarzyna Józwiak  
Koen K. Van de Vijver  
Marie-Jeanne T. F. D. Vrancken Peeters  
Frederieke van Duijnhoven  
Benno H. W. Hendriks  
Henricus J. C. M. Sterenborg  
Theo J. M. Ruers

# Influence of neoadjuvant chemotherapy on diffuse reflectance spectra of tissue in breast surgery specimens

Lisanne L. de Boer,<sup>a,\*</sup> Esther Kho,<sup>a</sup> Katarzyna Józwiak,<sup>b,c</sup> Koen K. Van de Vijver,<sup>d,e</sup> Marie-Jeanne T. F. D. Vrancken Peeters,<sup>a</sup> Frederieke van Duijnhoven,<sup>a</sup> Benno H. W. Hendriks,<sup>f,g</sup> Henricus J. C. M. Sterenberg,<sup>a,h</sup> and Theo J. M. Ruers<sup>a,i</sup>

<sup>a</sup>The Netherlands Cancer Institute, Department of Surgery, Amsterdam, The Netherlands

<sup>b</sup>The Netherlands Cancer Institute, Department of Epidemiology and Biostatistics, The Netherlands

<sup>c</sup>Institute of Biostatistics and Registry Research, Brandenburg Medical School Theodor Fontane, Neuruppin, Germany

<sup>d</sup>The Netherlands Cancer Institute, Department of Pathology, Amsterdam, The Netherlands

<sup>e</sup>Ghent University Hospital, Department of Pathology, Gent, Belgium

<sup>f</sup>Philips Research, Eindhoven, The Netherlands

<sup>g</sup>Delft University of Technology, Biomechanical Engineering Department, Delft, The Netherlands

<sup>h</sup>Amsterdam University Medical Center, Department of Biomedical Engineering and Physics, Amsterdam, The Netherlands

<sup>i</sup>University of Twente, TNW, Technical Medical Centre, Enschede, The Netherlands

**Abstract.** Diffuse reflectance spectroscopy (DRS) can discriminate different tissue types based on optical characteristics. Since this technology has the ability to detect tumor tissue, several groups have proposed to use DRS for margin assessment during breast-conserving surgery for breast cancer. Nowadays, an increasing number of patients with breast cancer are being treated by neoadjuvant chemotherapy. Limited research has been published on the influence of neoadjuvant chemotherapy on the optical characteristics of the tissue. Hence, it is unclear whether margin assessment based on DRS is feasible in this specific group of patients. We investigate whether there is an effect of neoadjuvant chemotherapy on optical measurements of breast tissue. To this end, DRS measurements were performed on 92 *ex-vivo* breast specimens from 92 patients, treated with neoadjuvant chemotherapy and without neoadjuvant chemotherapy. Generalized estimating equation (GEE) models were generated, comparing the measurements of patients with and without neoadjuvant chemotherapy in datasets of different tissue types using a significance level of 5%. As input for the GEE models, either the intensity at a specific wavelength or a fit parameter, derived from the spectrum, was used. In the evaluation of the intensity, no influence of neoadjuvant chemotherapy was found, since none of the wavelengths were significantly different between the measurements with and the measurements without neoadjuvant chemotherapy in any of the datasets. These results were confirmed by the analysis of the fit parameters, which showed a significant difference for the amount of collagen in only one dataset. All other fit parameters were not significant for any of the datasets. These findings may indicate that assessment of the resection margin with DRS is also feasible in the growing population of breast cancer patients who receive neoadjuvant chemotherapy. However, it is possible that we did not detect neoadjuvant chemotherapy effect in the some of the datasets due to the small number of measurements in those datasets. © The Authors. Published by SPIE under a Creative Commons Attribution 4.0 Unported License. Distribution or reproduction of this work in whole or in part requires full attribution of the original publication, including its DOI. [DOI: 10.1117/1.JBO.24.11.115004]

Keywords: neoadjuvant chemotherapy; diffuse reflectance spectroscopy; margin assessment.

Paper 190288R received Aug. 29, 2019; accepted for publication Nov. 4, 2019; published online Nov. 26, 2019.

## 1 Introduction

Diffuse reflectance spectroscopy (DRS) is an optical technology that measures the composition and morphology of tissue, based on absorption and scattering of light. A DRS measurement of tissue can be used to discriminate tissue types, as the optical characteristics differ between tissue types. The ability to discern healthy tissue from tumor tissue could potentially improve intra-operative margin evaluation during breast-conserving surgery and reduce the number of re-excisions, which are currently reported to be as high as 20% of patients.<sup>1–6</sup>

Previous research showed the potential of DRS to provide surgical guidance during breast-conserving surgery based on the ability of DRS to discriminate healthy breast tissue from tumor

tissue.<sup>7–11</sup> The reported accuracies for discriminating healthy breast tissue from tumor tissue reached up to 92% *in vivo*, albeit that these results were obtained in patients who did not have any “neoadjuvant chemotherapy.” In addition to the resection margin assessment, DRS technology has also been used for chemotherapy monitoring. Several studies indicate that the effect of chemotherapy can be measured with optical technologies.<sup>12–14</sup> The fact that the optical properties of tissue alter due to chemotherapy is not surprising, as effects of chemotherapy are seen in tumor tissue<sup>15–17</sup> as well as in healthy breast tissue.<sup>16–20</sup> However, currently, it is unclear if the contrast between healthy tissue and tumor tissue is affected by chemotherapy. It is important to consider the effect of chemotherapy on the DRS measurements, since there is an increasing tendency to treat breast cancer patients with chemotherapy before surgery as there are many benefits for the patient.<sup>21–26</sup> A preliminary study on the influence of neoadjuvant chemotherapy by our group did not

\*Address all correspondence to Lisanne L. de Boer, E-mail: [l.d.boer@nki.nl](mailto:l.d.boer@nki.nl)

find any significant differences between patients with and patients without neoadjuvant chemotherapy; however, the number of patients included in that study was limited ( $n = 9$ ), as well as the number of measurements per patient.<sup>27</sup>

Since our ultimate goal is to use the DRS technology during surgery for the detection of tumor deposits at the resection edge, it is essential to investigate if the changes induced by the neoadjuvant chemotherapy will affect the DRS measurements of the different tissue types. If the DRS measurements are profoundly different, then this might also affect the optical contrast between tumor tissue and healthy tissue types. This implies that, for the classification of DRS measurements of patients with neoadjuvant chemotherapy, the models based on the measurements of patients without neoadjuvant chemotherapy are not suitable. The goal of our study, therefore, is to assess whether the spectrum of various tissue types, for example, pure “fat” or a combination of fat and “connective” tissue, are significantly different in patients with and without neoadjuvant chemotherapy. In other words, we evaluate whether neoadjuvant chemotherapy induces changes in the breast tissue which may affect the use of DRS for margin detection.

We evaluate the optical measurements of surgical breast cancer specimens in a relatively large dataset of patients with and without neoadjuvant chemotherapy, when compared with other studies. First, we investigate the difference in the tissue composition between patients with and without neoadjuvant chemotherapy. For this purpose, we compare the histopathology of both the whole tissue slice and the measurement locations. Subsequently, two analyses were performed. In the first analysis, we compared if the measured diffuse reflectance at any of the wavelengths was significantly different in patients with or without neoadjuvant chemotherapy (spectral analysis). For the second analysis, fit parameters were derived from the measured spectra using an analytical fit model to quantify the measured spectra. These fit parameters were then compared to assess whether these were significantly different for the different tissue types in patients with and without neoadjuvant chemotherapy (fit parameter analysis).

## 2 Materials and Methods

### 2.1 DRS Measurements

#### 2.1.1 Specimens

All measurements were performed on *ex-vivo* breast samples from either breast-conserving surgeries or mastectomy procedures. No written informed consent from patients was required according to Dutch law (WMO) for the measurements. There was no selection of specimens based on tumor grade or type of breast cancer. Fresh specimens were brought to the pathology department where, after inking of the margins and slicing the specimen in a bread-loafed manner, one slice was provided for the optical measurements. Before measuring, the tissue was placed on top of a piece of black rubber in a macrocassette. All tissue slices were at least 2-mm thick.

#### 2.1.2 Optical measurements

DRS measurements were obtained between 400 and 1600 nm. Details on the measurement setup were reported previously.<sup>28,29</sup> In short, the light of a halogen light source was transferred to the tissue by means of the illuminating fiber that is integrated into a fiber-optic probe. Two other integrated fibers were attached to

two spectrometers, one covering the visual wavelength range (Andor Technology, DU420A-BRDD, 400 to 1000 nm) and one covering the near-infrared wavelength range (Andor Technology, DU492A-1.7, 900 to 1600 nm), both of which collected the photons after interacting with the tissue. The setup was controlled with LabVIEW® (Austin, Texas) software that performed a calibration from photon counts to diffuse reflectance and combined the output of both spectrographs to form one continuous spectrum from 400 to 1600 nm. For the calibration, a cap with Spectralon® (SRT-99-100, Labsphere, Inc., Northern Sutton, New Hampshire) at the bottom was used. The fiber distance between the emitting and collecting fiber was 1 mm.

#### 2.1.3 Grid

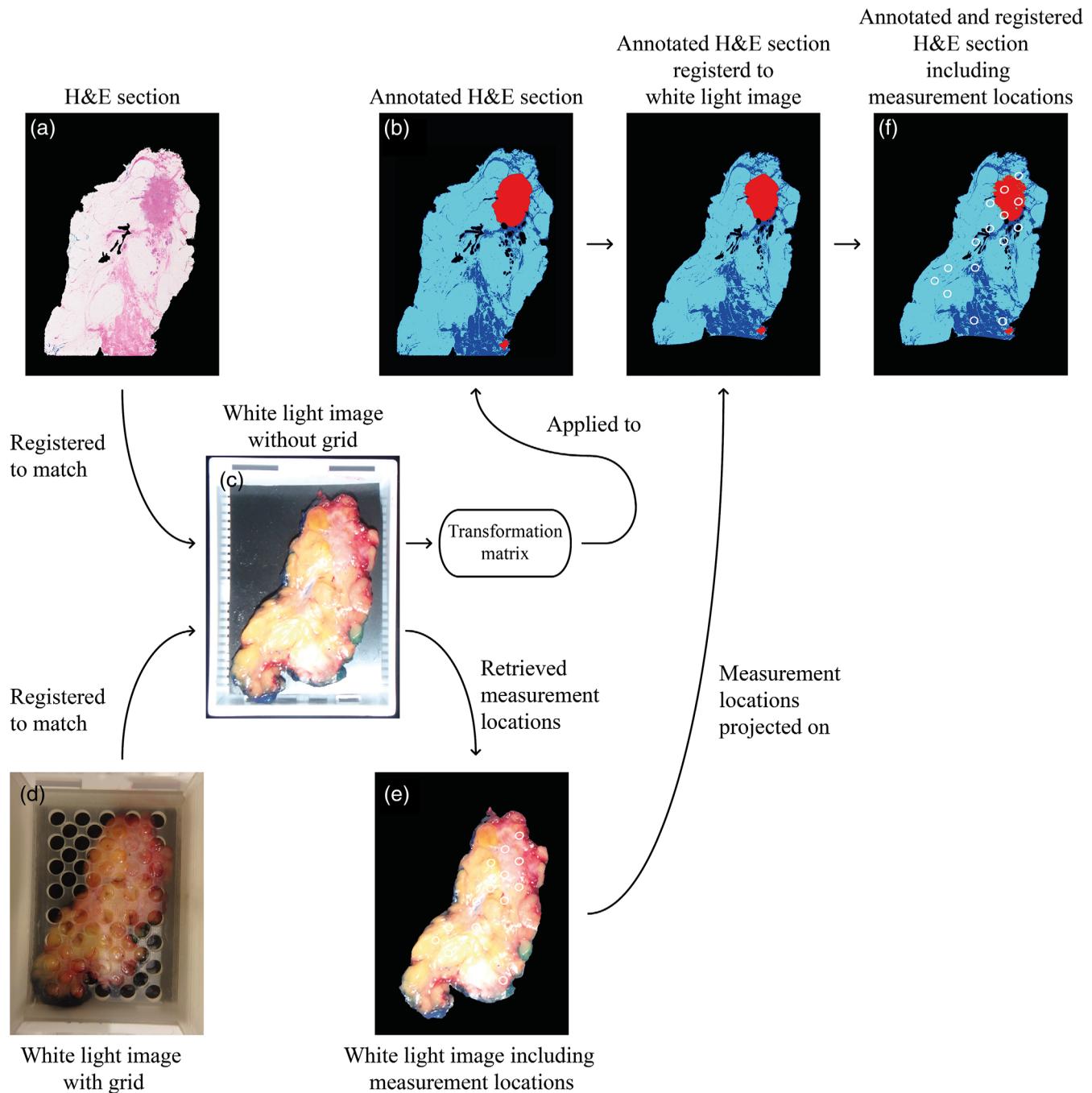
During the measurements, a custom-made grid was used to ensure a robust correlation between the locations of the optical measurements and the corresponding histopathology. Before the fiber-optic measurements, two white light images were acquired of the tissue in the cassette (1) with the grid and (2) without the grid. After the optical measurements, the grid was removed and tissue was processed according to standard protocol resulting in a hematoxylin and eosin (H&E) stained section of the whole tissue slice.

### 2.2 Correlating DRS Measurement to Histopathology

The whole H&E section [Fig. 1(a)] was evaluated by a pathologist who annotated areas with invasive carcinoma, ductal carcinoma *in situ* (DCIS), and glandular ducts. The section and annotations of the pathologist were digitalized and, with thresholding, all other pixels in the H&E image were labeled as fat or connective in MATLAB R2018a, based on the fact that fat areas appear white in the H&E section, whereas connective tissue will stain pink. Glandular tissue was included in the connective tissue class. The result was an image in which each pixel had a label corresponding to a tissue type (fat, connective, or invasive carcinoma) [Fig. 1(b)]. The histopathology was registered with the white light image without the grid [Fig. 1(c)] to adjust for tissue deformation, which was introduced during histopathological processing. Another registration was made between this white light image and another white light image including the grid [Fig. 1(d)] to retrieve the actual measurement locations in the white light image [Fig. 1(e)]. Combining both registrations then allowed projecting the measurement locations in the annotated H&E section, which was corrected for tissue deformation [Fig. 1(f)]. This registration method is described in more detail in a previous paper.<sup>30</sup> To calculate the mean percentage of tumor cells, an estimation was made of the ratio between tumor cells and connective tissue in all areas that were considered invasive carcinoma. As there were few measurement locations with DCIS ( $n = 4$ ) in the chemotherapy patient group and the percentage of DCIS in those locations was less than 20%, this tissue type was not included in the analysis.

### 2.3 Composition of Measurement Locations

To investigate if the locations that were measured with the fiber-optic probe were a good representation of the tissue composition of the specimens, the composition of the tissue in the whole H&E sections of the specimens (from patients with and

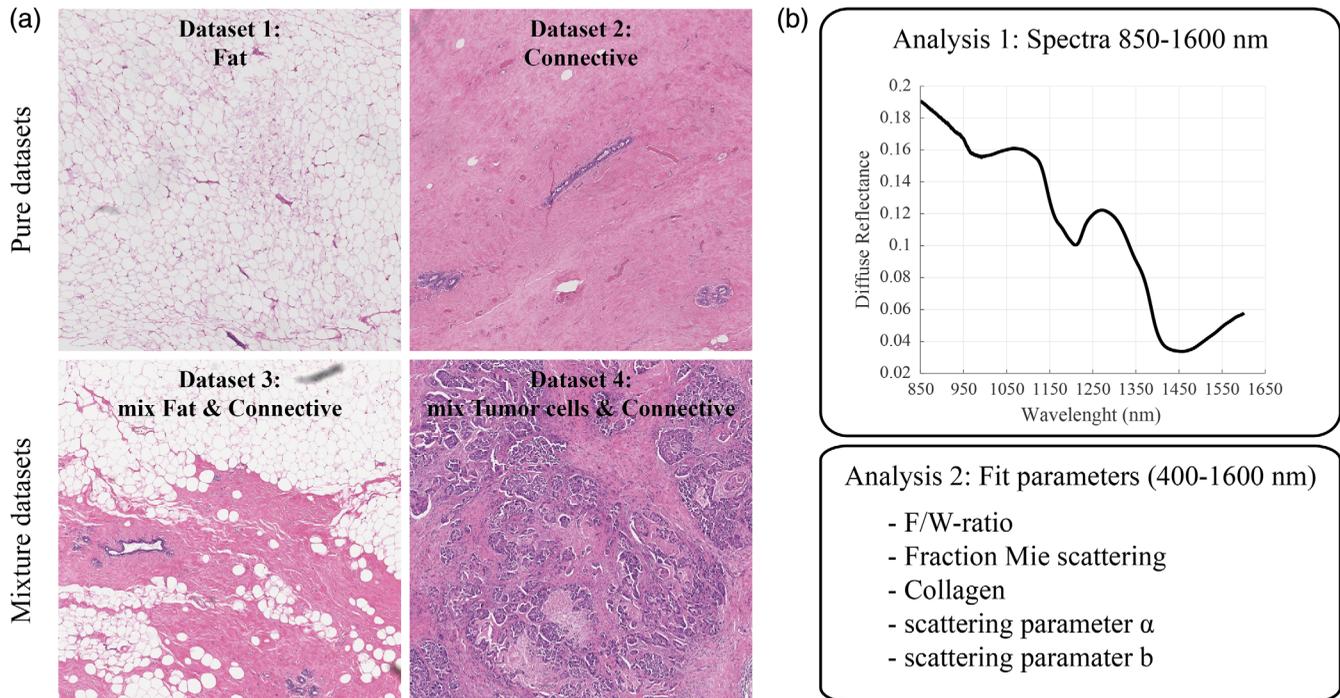


**Fig. 1** Schematic overview of method for correlating the histopathology with the optical measurement locations. A registration is made between (a) the H&E section and (c) the white light without the grid. The transformation matrix obtained after this registration procedure is applied to (b) the H&E section with annotations. This way the annotated image is corrected for deformations in the H&E section induced during the processing of the tissue at the pathology department. Using (d) a white light image with the grid, the measurement locations are retrieved. (e) The retrieved measurement locations can be projected on the H&E section (as both are registered to the white light image). (f) The end result is an image including the measurement locations that is corrected for tissue deformation, and in which each pixel is annotated.

without neoadjuvant chemotherapy) was compared with the composition of the measurement locations (from patients with and without neoadjuvant chemotherapy). A two-sided  $t$ -test was used to evaluate if the percentages of fat, connective, and tumor cells in the locations measured with the fiber-optic probe were significantly different from the percentages of those tissue types present in the whole section of the specimen. A  $p$ -value of 0.05 or smaller was considered significant.

## 2.4 Datasets and Analysis

In the analysis, we used four datasets comprising measurements with the same histopathology (Fig. 2). Datasets 1 and 2 consisted of locations that were  $>95\%$  fat tissue or  $>95\%$  connective tissue, respectively. Dataset 3 consisted of measurement locations that were a mixture of fat and connective tissues. The last dataset, dataset 4, comprised all locations that contained



**Fig. 2** (a) Examples of histopathology in the datasets used in the analyses. Locations were selected from the specimens of patient with and without chemotherapy, which consisted of >95% fat tissue (dataset 1), >95% connective tissue (dataset 2), a mixture of fat and connective tissue (dataset 3), or a mixture of tumor cells and connective tissue (dataset 4). (b) With each dataset, two analyses were performed, either based on the spectra (850 to 1600 nm) or fit parameters, in which the measurements of patient with and without chemotherapy are compared.

tumor cells in combination with some connective tissues. To select the measurements for each dataset, the estimated percentages of fat, connective, and tumor cells were used, as described in Sec. 2.2. In each of the four datasets, the difference between measurements with and without neoadjuvant chemotherapy was assessed by performing two analyses.

#### 2.4.1 Spectral analysis

For the first analysis, only the intensities measured between 850 and 1600 nm were used. Although the measurements were acquired over the wavelengths between 400 and 1600 nm, the part of the spectrum between 400 and 850 nm was excluded in the analysis as pathology ink, present on the specimens, strongly influenced the measured intensities of the wavelengths below 850 nm. The spectra were not normalized before the analysis.

#### 2.4.2 Fit parameter analysis

For the second analysis, the fit parameter analysis, the spectra were fitted with an analytical fit model, which was described in previous publications.<sup>28,29</sup> This model estimated the composition of the measurement volumes based on diffusion theory using the measured spectrum and the absorption spectra of substances known to be present in the tissue. Included in the model were oxygenated and deoxygenated hemoglobin, fat, water, and collagen. The fit parameters, derived from the spectra with the analytical fit model, which are used in the analysis were: ratio between fat and water (F/W-ratio), fraction Mie scattering, Collagen,  $\alpha$ , and  $b$ . The latter two define the amplitude ( $\alpha$ ) and slope ( $b$ ) in the function that describes the reduced scattering.<sup>28</sup>

To account for the influence of the pathology ink, spectra of the pathology inks were also included as fit parameters.

## 2.5 Statistics

In both analyses, the datasets were evaluated one by one via generalized estimating equation (GEE) models. This statistical approach is specifically suitable for modeling correlated data, such as repeated measurements in one patient or repeated measurements of tissue types in multiple patients. In such a case, incorporating within-subject and between-subject variations in the statistical analysis is important.<sup>31</sup> A GEE model evaluates the association between measurements and covariates while taking into account the possible relations between measurements that are obtained in one patient.<sup>32</sup> In the GEE models, an equicorrelated structure was used to describe the variance and covariance between the repeated measurements.

In the first analysis, a GEE model was generated for each wavelength, whereas in the second analysis a GEE model was generated for each fit parameter. Covariates included in the models were neoadjuvant chemotherapy (“yes”/“no”) and “menopausal status” (“premenopausal or perimenopausal”/“postmenopausal”). Menopausal status was incorporated in the models, as this factor is known to be related to breast density.<sup>33,34</sup>

Datasets 3 and 4 contained measurements that were a mixture of connective tissues in combination with either fat tissues or tumor cells. Since the ratio between two tissue types can also affect the spectrum, the percentage connective tissue estimated from the annotated H&E section was added as an extra covariate

in the GEE models of these datasets to take into account this effect.

The analyses were performed in IBM SPSS Statistics 25 with help of the SPSS Python plugin module that provided the possibility to iterate over either the wavelengths or the fit parameters, making a GEE model for each one of them. The  $\alpha$ -level was set to 0.05 for all the GEE models.

### 3 Results

In this study, measurements from 92 patients were used, of which 30 patients were treated with neoadjuvant chemotherapy. From each patient, one specimen was used for the optical measurements. The patient characteristics are listed in Table 1. The mean age of the group of pretreated patients was slightly younger, compared to the mean age of the group of patients without neoadjuvant chemotherapy. Furthermore, the majority of patients who were treated with neoadjuvant chemotherapy received a combination of different types of chemotherapeutics (anthracycline, alkylating agents, taxanes, and antimetabolites).

The response to neoadjuvant chemotherapy was evaluated by (1) comparing the tumor size on magnetic resonance imaging pretreatment and post-treatment (radiological response) and (2) histopathological assessment of the pathologist (pathological response). The combination of the radiological and pathological responses is also listed in Table 1. In 12 patients (40%), the response to neoadjuvant chemotherapy was complete, according to both the radiological and the pathological assessments. In another six patients (20%), both evaluation methods found the response to be “partial.” If a favorable or minimal radiological response is also considered as partial response, in five more patients (16.7%) the response in both evaluation methods is partial. In one patient, the radiological evaluation reported a progressive disease and the pathologist noted no response in the pathology report.

In five patients, either the radiological or pathological response was complete, whereas the other evaluation method asserted a partial response. In one patient, the radiological response was considered partial, whereas the pathologist stated that there was no response. Thus, in six patients (20%), the radiological response was not in accordance with the pathological response.

#### 3.1 Composition of the Tissue Section

Figure 3 shows the mean percentages and standard deviations of the percentage of fat, connective tissue, and tumor cells in the whole H&E section and the measurement locations measured with the fiber-optic probe, from patients without neoadjuvant chemotherapy [Fig. 3(a)] and with neoadjuvant chemotherapy [Fig. 3(b)]. In the data from patients without neoadjuvant chemotherapy, the mean percentages of fat, connective tissue, and tumor cells calculated in the whole section are 63%, 40%, and 5%, respectively. For these tissue types in the same order, the mean percentages in the measured locations were 58%, 33%, and 6%, respectively. With a *t*-test, we did not find evidence of differences in the percentages of fat, connective tissue, and tumor cells between the whole sections and the measurement locations. None of the *p*-values were significant (fat: 0.30, connective tissue: 0.10, and tumor cells: 0.63).

The calculated mean percentages of fat, connective tissue, and tumor cells in the whole sections from patients with neoadjuvant chemotherapy were 61%, 39%, and 2%, whereas in the measured locations these mean percentages were 58%, 41%, and 2%, respectively [Fig. 3(b)]. Similar to the data from

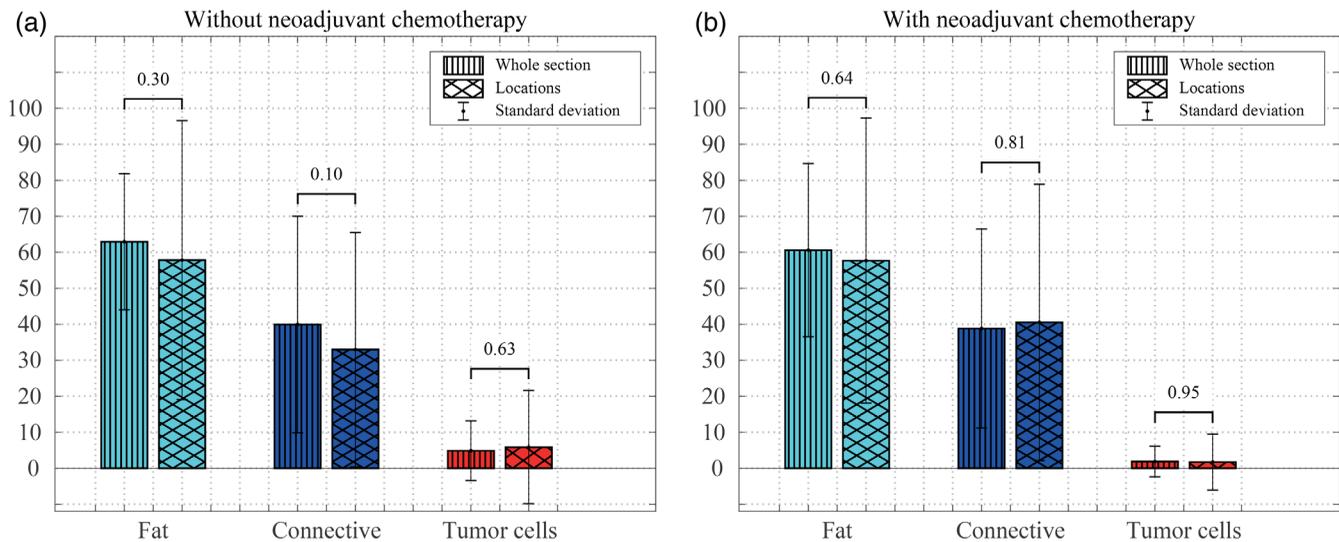
**Table 1** Patient characteristics of specimens that were used in the analysis.

	No chemo ( <i>n</i> = 62)	Chemo ( <i>n</i> = 30)
<b>Age (std)</b>	58 ( $\pm$ 10.6)	51 ( $\pm$ 11.5)
<b>Menopausal status</b>		
Premenopausal	22 (36.1%)	13 (43.3%)
Perimenopausal	5 (8.2%)	3 (10%)
Postmenopausal	34 (55.7%)	14 (46.7%)
<b>Chemotherapy</b>		
AC	—	6 (20%)
AC + P	—	7 (23.2%)
AC + CP + P	—	3 (10%)
AC + P + C	—	1 (3.3%)
AC + CP	—	1 (3.3%)
AC + T	—	1 (3.3%)
AC + CP + P + C	—	1 (3.3%)
PTC	—	5 (16.7%)
FEC + PTC	—	1 (3.3%)
TAC	—	1 (3.3%)
P	—	2 (6.6%)
AC + P + CP + CAP	—	1 (3.3%)
<b>Response</b>		
Radiological complete response and pathological complete response <sup>a</sup>	—	12 (40%)
Radiological complete response and pathological partial response <sup>b</sup>	—	4 (13%)
Radiological partial response and pathological complete response <sup>b</sup>	—	1 (3.3%)
Radiological partial response and pathological partial response <sup>a</sup>	—	6 (20%)
Radiological partial response and no pathological response <sup>b</sup>	—	1 (3.3%)
Favorable radiological response and partial pathological response <sup>a</sup>	—	4 (13%)
Minimal radiological response and partial pathological response <sup>a</sup>	—	1 (3.3%)
Progressive disease (radiology) and no pathological response <sup>a</sup>	—	1 (3.3%)

A, adimycin; C, cyclophosphamide; P, paclitaxel; CP, carboplatin; T, docetaxel; F, 5-fluorouracil; E, epirubicin; CAP, capecitabine.

<sup>a</sup>Radiological and pathological responses in accordance.

<sup>b</sup>Radiological and pathological responses not in accordance.



**Fig. 3** Composition of H&E section and the measurement locations from specimens of patients (a) without chemotherapy and (b) with chemotherapy. The bars with the vertical stripes represent the mean percentages of these tissue types calculated in the whole tissue section. The bars with the crossed pattern represent the mean percentages of the tissue types calculated in the measurement locations. The error bars show the standard deviation. The numbers above the bars belonging to one tissue type display the results of the *t*-test comparing the percentages measured in the whole section and the percentages in the measured locations.

the patients without chemotherapy, a *t*-test was used to assess if the percentages were significantly different in the whole sections compared with the measured locations. The *p*-values for fat, connective tissue, and tumor cells were 0.64, 0.81 and 0.95, respectively, and thus not significant.

It was concluded that there was no bias in the selection of the fiber-optic probe measurement locations and that this dataset can be seen as a good representation of the tissue that is present in the whole section.

Table 2 lists the number of measurements with and without neoadjuvant chemotherapy in each of the four datasets, and the number of patients the measurements originated from, as well as the mean number of measurements per specimen with the standard deviation. The datasets including measurements of pure connective tissue (dataset 2) and measurements of a mixture of connective tissue and tumor cells (dataset 4) are relatively small and originate from a limited number of patients. Although the mean percentage of connective tissue is rather high (Fig. 3), locations of >95% connective tissue are quite rare. More often

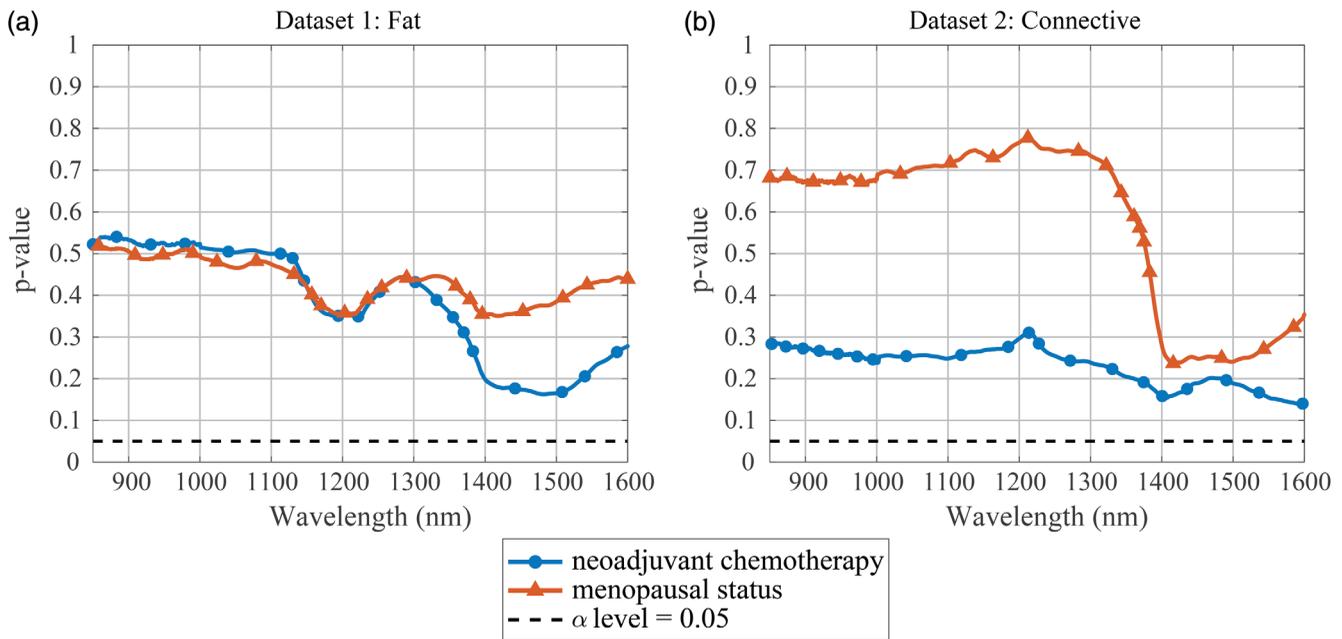
connective tissue is present in combination with fat tissue (dataset 3).

### 3.2 Intensity Measured between 850 and 1600 nm (Analysis 1)

With the GEE models, for each wavelength, a *p*-value was calculated to assess if the difference between the measured intensities in two tissue types was significant. The results of the GEE models for the pure datasets (pure fat and pure connective tissue) are shown in Fig. 4. As shown in Figs. 4(a) and 4(b), none of the wavelengths reach a *p*-value of 0.05 or lower. Thus, for pure fat or pure connective tissue, there are no wavelengths that have a significantly different intensity due to chemotherapy. Thus, no evidence was found for rejecting our null hypothesis that chemotherapy had no effect on the measurement. Furthermore, between premenopausal/perimenopausal and postmenopausal women no significant difference is seen. It should be noted, however, that for the pure connective

**Table 2** Datasets used in the two analyses.

	No neoadjuvant chemotherapy		With neoadjuvant chemotherapy	
	Total measurement locations (# of patients)	Average of measurement locations/patient (std)	Total measurement locations (# of patients)	Average of measurement locations/patient (std)
Dataset 1: Fat	222 (49)	4.5 (4.0)	144 (24)	6 (4.7)
Dataset 2: Connective tissue	19 (13)	1.5 (0.8)	32 (9)	3.6 (2.1)
Dataset 3: Fat/connective tissue	316 (56)	5.6 (4.5)	236 (30)	7.9 (4.7)
Dataset 4: Tumor cells/connective tissue	43 (21)	2.0 (1.3)	13 (6)	2.2 (1.5)
<b>Total # measurements</b>	<b>600</b>		<b>425</b>	

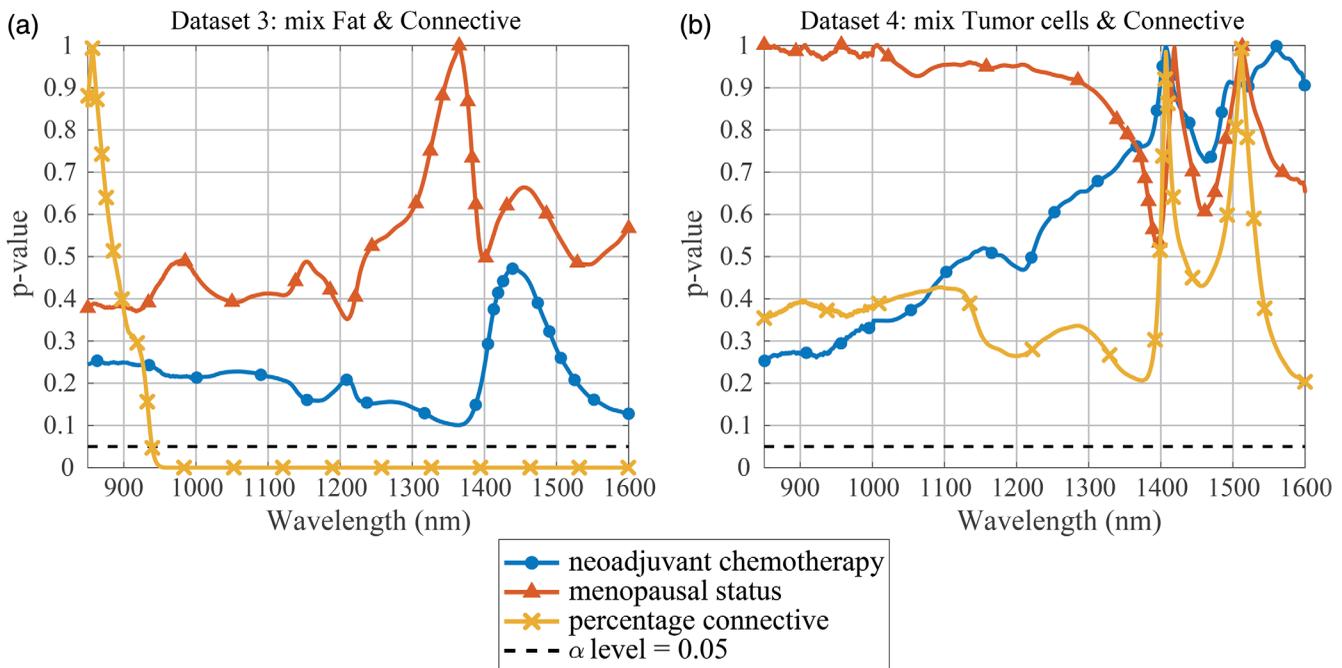


**Fig. 4** Result of GEE models: (a) *p*-values for pure fat (dataset 1) and (b) pure connective tissue (dataset 2). None of the wavelengths are significant for either two datasets comparing the intensities measured in patients with and without chemotherapy or comparing postmenopausal patients with premenopausal/ perimenopausal patients.

measurements a limited number of data points were available, which cause no detection of an existing effect.

In Fig. 5, the results of the GEE models for the mixture datasets are displayed. Figure 5(a) shows the results of the GEE

models for the measurements of connective tissue in combination with fat tissue. There was no evidence found for rejection of our null hypotheses since a significant difference in intensity measured at any of the wavelengths in the group with



**Fig. 5** Result of GEE models in (a) *p*-values for the two mixture datasets, with connective tissue in combination with fat tissue (dataset 3) and (b) tumor cells (dataset 4). In the results of the dataset consisting of measurement locations that were a mixture of fat and connective tissue (dataset 3), chemotherapy and menopausal status were not significantly different for any of the wavelengths. The percentage connective tissue in the measurement location was significant for wavelengths above 950 nm. The results of the dataset that consisted of measurement locations with tumor cells and connective tissue (dataset 4) are displayed in (b). Here, neither chemotherapy, nor menopausal status, nor the percentage connective tissue in the measurement location resulted in a significantly different measured intensity.

neoadjuvant chemotherapy compared with the group without neoadjuvant chemotherapy was detected. Neither did the menopausal status of the patient result in a difference in intensity measured for any of the wavelengths. As explained in Sec. 2, in the GEE models of the mixture datasets, the percentage of connective tissue was added as an extra covariate. In the dataset including a mixture of fat and connective tissue, covariate describing the percentage of connective tissue in the measurement location was significant for the wavelengths above 950 nm. In other words, the intensity measured at these wavelengths is dependent on the amount of connective tissue in the measurement.

In the other mixture dataset, with measurement locations that included tumor cells in combination with connective tissue (dataset 4), neither neoadjuvant chemotherapy nor the menopausal status showed significant effect on the intensity at any of the wavelengths [Fig. 5(b)]. In contrast to the other mixed tissue data set (dataset 3), the percentage connective tissue, or vice versa, in this case, the percentage of tumor cells, did not result in a significantly different intensity at any of the wavelengths.

### 3.3 Fit Parameters (Analysis 2)

For each of the fit parameters that were deduced from the DRS spectrum, a GEE model was generated for each dataset. The

**Table 3** Result in  $p$ -values of the GEE models of fit parameters of the datasets of fat tissue (dataset 1) and connective tissue (dataset 2).

Fit parameter	Fat dataset (dataset 1)		Connective tissue dataset (dataset 2)	
	Neoadjuvant chemo	Menopausal status	Neoadjuvant chemo	Menopausal status
F/W ratio	0.51	0.43	0.19	0.77
Fraction Mie scattering	0.52	0.99	0.48	0.82
Collagen ( $\mu$ M)	0.07	0.73	0.61	0.24
$\alpha$	0.49	0.19	0.12	0.94
b	0.88	0.63	1.00	0.66

**Table 4** Result in  $p$ -values of the analysis of fit parameters for the mixture datasets of a combination of connective tissue with fat or tumor cells (datasets 3 and 4).

Fit parameter	Fat and connective tissue (dataset 3)			Tumor cells and connective tissue (dataset 4)		
	Neoadjuvant chemo	Menopausal status	% connective tissue	Neoadjuvant chemo	Menopausal status	% connective tissue
F/W ratio	0.21	0.41	0.00*	0.11	0.40	0.36
Fraction Mie scattering	0.13	0.22	0.05*	0.64	0.32	0.39
Collagen ( $\mu$ M)	0.26	0.48	0.00*	0.00*	0.01*	0.03*
$\alpha$	0.56	0.10	0.00*	0.87	0.32	0.34
b	0.44	0.08	0.00*	0.91	0.28	0.99

\* = Significant.

$p$ -values obtained for each of the fit parameters from the dataset with fat measurements (dataset 1) and the dataset composed of connective tissue measurements (dataset 2) are listed in Table 3.

In the fat measurements (dataset 1), none of the fit parameters were significantly different due to neoadjuvant chemotherapy or menopausal status of the patient. As for the connective tissue dataset (dataset 2), similarly, no significant differences were found for any of the fit parameters between the group of patients with or without neoadjuvant chemotherapy and between the groups of patients who were premenopausal/perimenopausal or postmenopausal. This suggests that there is no evidence for rejecting the null hypothesis of no effect of the chemotherapy or menopausal status.

Table 4 includes the  $p$ -values for the mixture datasets, in which connective tissue is mixed with either fat tissue (dataset 3) or tumor cells (dataset 4). In the analysis of dataset 3, none of the fit parameters were significantly different, when comparing the measurements of patients with neoadjuvant chemotherapy with the measurements of the patients without neoadjuvant chemotherapy. Also, the menopausal status of the patient did not have a significant influence on any of the five fit parameters. The percentage of connective tissue was highly significant for all fit parameters, except for the fraction Mie scattering, which was borderline significant. It seems that the amount of connective tissue in the measurement location had a clear effect on the fit parameters.

The results of the analysis of dataset 4 do not show enough evidence for a significant influence of neoadjuvant chemotherapy and menopausal status on four of the five fit parameters (F/W ratio, fraction Mie scattering,  $\alpha$ , and b). Also, the amount of connective tissue did not show a sign of influence on these fit parameters. Only the fit parameter collagen was significantly different for all covariates, suggesting that in this case there is some evidence of a possible relationship between the measurement of this fit parameter, the preoperative treatment, and menopausal status of the patient and the percentage of connective tissue in the measurement volume.

## 4 Discussion

This research was focused on investigating whether DRS measurements of breast tissue are affected by neoadjuvant chemotherapy. First, the composition of tissue types of the whole H&E sections from patients with and without neoadjuvant chemotherapy was analyzed, as well as the composition of the

locations that were measured with the fiber-optic probe. The compositions were compared with investigate if the measured locations were representative of the tissue types in the specimen as seen in the whole H&E section. Next, the DRS measurements of the *ex-vivo* specimens of patients with and without neoadjuvant chemotherapy were compared by analyzing both the diffuse reflectance intensity data over the spectrum between 850 and 1600 nm (analysis 1) and the fit parameter data derived from the spectrum between 400 and 1600 nm (analysis 2). Both analyses were performed with GEE models that are suitable for assessing correlated data. In the GEE models, in addition to treatment with neoadjuvant chemotherapy, other possible confounding factors were included, such as the menopausal status, and for the mixture datasets, the amount of connective tissue, as these could also affect the DRS measurement.

Comparison of the percentages of fat, connective tissue, and tumor tissue present in the fiber-optic probe measurement locations with the percentages in the whole H&E section showed that these were not significantly different. Thus, showing that the probe measurements (which in essence are a subset of the whole sections) may be a representation of the tissue in both the patient groups, without and with neoadjuvant chemotherapy.

Subsequently, the intensity data (analysis 1: spectral analysis) was used to assess if there were wavelengths with significant different intensity in the patients treated with neoadjuvant chemotherapy and the patients that received treatment without neoadjuvant chemotherapy. This was done in four separate datasets, in which all measurements in one dataset had similar histopathology. Significance was not detected in all four datasets based on the measured intensity, indicating no intensity differences related to neoadjuvant chemotherapy at any of the wavelengths from 850 to 1600 nm. In the analysis, the menopausal status also did not show a significant result for any of the wavelengths in any of the datasets.

In the mixture datasets, consisting of connective tissue in combination with fat (dataset 3) or tumor cells (dataset 4), the percentage of connective tissue in the measurement was included as a covariate. In the dataset that included connective tissue with fat (dataset 3), this covariate was significant for wavelengths above 950 nm, indicating that the measured intensity at these wavelengths was significantly different in these two tissue types [Fig. 5(a)]. This is in line with the expectations, as there are clear biological differences between fat tissue and connective tissue that are displayed in the higher wavelength range. However, in the other dataset in which connective tissue was mixed with tumor cells, none of the wavelengths were significant. This finding may suggest that discriminating connective tissue from tumor cells is more difficult than discriminating connective tissue from fat tissue. For discriminating fat from connective tissue, the observed wavelengths can be used. For discriminating connective tissue from tumor cells, no such clear individual wavelengths could be identified by this approach, probably due to the fact that connective/glandular tissue can resemble tumor tissue.<sup>35,36</sup> Potentially, a combination of wavelengths that are individually not significant could discriminate connective tissue from tumor cells, but this is not investigated in the current analysis.

The fit parameters that were derived from the measured spectra by using an analytical fit model were also evaluated with GEE models (analysis 2). As for the datasets of pure fat tissue and pure connective tissue (datasets 1 and 2), none of the fit parameters were significantly different in the groups with or

without neoadjuvant chemotherapy and the groups of premenopausal/perimenopausal or postmenopausal patients. For these datasets, the results of analyses 1 and 2 are the same.

In the analysis of the mixture dataset that consisted of a combination of fat tissue and connective tissue (dataset 3), the fit parameters were also not significantly different in the measurements of patients who did or did not have neoadjuvant chemotherapy treatment or patients who were premenopausal/perimenopausal or postmenopausal. The percentage of connective tissue was significant for all fit parameters, indicating that all fit parameters changed significantly if the percentage of connective tissue changed and can thus contribute to discriminating fat tissue from connective tissue. For this dataset, the outcomes of analyses 1 and 2 are again the same. This is also in line with the expectations as the absorption coefficient of collagen, the predominant tissue constituent present in connective tissue, is different from the absorption coefficient of fat.<sup>37,38</sup>

As for the last dataset, with measurements of tumor cells and connective tissue, four of the five fit parameters were not significant for any of the covariates, confirming the results of the first analysis performed with the measured intensities. These results are in line with the results of the spectral analysis, in which no significance was found for neoadjuvant chemotherapy, menopausal status, and connective tissue content. However, the fit parameter representing the amount of collagen in a measurement location appeared significant for all three covariates (neoadjuvant chemotherapy, menopausal status, and percentage of connective tissue). The fact that the fit parameter that describes the amount of collagen was related to the treatment with neoadjuvant chemotherapy, as well as menopausal status, as well as the percentage of connective tissue, may suggest that this fit parameter is significantly different in measurements of patients with and without chemotherapy, in patients with different menopausal status, as well as in measurements with different percentage of connective tissue. It was, therefore, concluded that this fit parameter should be used with caution when used for discriminating healthy tissue from tumor tissue in a dataset that includes patients with and without chemotherapy and with different menopausal statuses.

To our knowledge, this is the first research that assessed the effect of neoadjuvant chemotherapy on the optical spectra measured with DRS using a relatively large number of specimens. Nevertheless, the datasets of measurement locations of pure connective tissue and connective tissue in combination with tumor cells are small and, thus, conclusions should be drawn with reservation. The absence of evidence for significant differences in these datasets might be related to the small number of measurements.

There are a number of publications that describe the microscopic changes that were observed by the pathologist under the microscope or on magnetic resonance imaging. Commonly seen phenomena in nontumor-bearing breast tissue include lobular atrophy and atypia, as well as lobular sclerosis. We did not find any difference in the healthy measurements, which might be explained by the fact that the described structures are relatively small in comparison to our measurement volume. Some studies also report on a decrease in breast density. However, this effect was more profound in younger patients (<40 years).<sup>18</sup> The mean ages of the patients to whom the specimens in this study belonged were 58 (without chemotherapy) and 51 (with chemotherapy) years. Potentially the involution of breast tissue was already present in these patients and, therefore, little difference

is seen between the ratio of fat tissue and connective tissue (Fig. 3). Another explanation could be that the morphological changes that are seen as a decrease in breast tissue are present on a larger scale (centimeters) than the fiber distance used in this study (>1 mm) and were, therefore, not detected by the DRS measurements. In tumor tissue, the decrease in cellularity and formation of fibrous areas are often described as a marked feature of chemotherapy treatment. The reduction in the number of tumor cells and the increase in fibrosis could be an explanation for the difference found in the fit parameter that expresses the amount of collagen. However, it should be noted that including the percentage of tumor cells in the GEE model as a covariate should account for this, unless the estimations of the percentage tumor cells based on the H&E sections are not representative of the true percentages of tumor cells in the actual measurement volume.

Diffuse optical spectroscopy has been researched as clinical tool for monitoring chemotherapy treatment and even for predicting therapy response.<sup>24,39-42</sup> However, the main focus of these published papers is on comparing pre- and post-chemotherapy measurements of individual patients, whereas we are interested in comparing groups of patients with and without neoadjuvant chemotherapy. The results presented in this paper show no evidence for a profound effect of chemotherapy on the optical measurements. This does certainly not imply that chemotherapy response monitoring is not possible using DRS. It merely means that the contrast between healthy tissue and tumor tissue is not altered as a consequence of the neoadjuvant chemotherapy. In other words, the composition of the tissue might change due to treatment with neoadjuvant chemotherapy, but the optical response of the tissue type in itself is not significantly different in patients with and without neoadjuvant chemotherapy.

A previous study by Laughney et al.,<sup>43</sup> designed to investigate the use of spatial frequency domain imaging for margin assessment of breast specimens, also included patients who were treated with neoadjuvant chemotherapy and assessed the influence of chemotherapy on the optical contrast between tumor and healthy tissues. It was reported that measurements of tumor tissue that had been treated with neoadjuvant chemotherapy had an increased scattering slope, compared with the tumor measurements of patients without neoadjuvant chemotherapy, which was suggested to be related to cell shrinkage. This is different from our results as no difference was found for the scattering parameters. In another study by Keller et al.,<sup>44</sup> diffuse reflectance spectra of patients with and without neoadjuvant chemotherapy were measured and compared. They were able to discriminate normal measurements of patients who received neoadjuvant chemotherapy from normal measurements of patients without treatment with neoadjuvant chemotherapy, suggesting that there might be an influence of chemotherapy. However, in both studies, the analysis was based on a very limited number of patients ( $n = 5$  and  $n = 3$ , respectively).

The response according to pathology and/or radiology was not included as a covariate in the GEE models, as the definition of response can be difficult and subjective.<sup>15</sup> This can also be deduced from Table 1, which shows that in six patients (20%) the radiological response is different from the pathological response. Furthermore, patients were not subdivided based on neoadjuvant chemotherapy regime since patients are routinely treated with a combination of chemotherapeutics that have different mechanisms of action and thus all types of reactions are potentially present in each patient. In this study, we did not

consider patients who were treated with neoadjuvant hormonal therapy. From the literature, it is known that this type of therapy can also induce tissue changes.<sup>45-47</sup> However, the number of patients who were treated with neoadjuvant hormonal therapy during the time of this study was too small to include this group of patients in the analyses.

Since measurements were taken from *ex-vivo* samples after some processing at the pathology department, there was contamination of pathology ink in the measurements. The ink was deposited on the tissue slices by the knife when the tissue was sliced and by the gloves of the pathologist when palpating the tissue slices. By excluding all wavelengths below 850 nm and fit parameters of this wavelength region, influence of pathology ink was reduced as much as possible. However, as a consequence, the impact of chemotherapy on the visual wavelength range could not be investigated. In addition, with the current setup, we did not investigate if there are differences in these tissue types if measurements were acquired *in vivo*. However, it is not expected that the wavelength range that was evaluated in this study is different in the *in-vivo* setting compared with the *ex-vivo* setting.<sup>27</sup> The ultimate goal is the intraoperative application of the technology, allowing the surgeon to assess the tissue when performing the resection. Future research, therefore, is directed toward *in-vivo* studies, which allow acquiring data from this visual part of the spectrum without the contamination of the pathology ink. *In-vivo* application of the technology also allows acquiring DRS measurements with larger fiber distances (~3 mm) to increase the depth sensitivity of the system. In this study, this was not possible as the thickness of the tissue was a constraint in the maximum fiber distance that could be used.

In conclusion, we used two different analyses (diffuse reflectance intensity at a specific wavelength and fit parameter data) to assess the influence of chemotherapy on optical characteristics of breast tissue by assessing four different datasets in which all measurements in one dataset had similar histopathology. In addition, the influence of menopausal status on the measurements was assessed, and for the datasets with a mix of tissue types, the percentage of connective tissue was included in the analysis. The measured intensity for the different tissue types was not significantly different in patients with or without neoadjuvant chemotherapy in any of the four datasets. Also, comparing the intensity in the measurements of the group of patients who were postmenopausal to those who were premenopausal/perimenopausal, no significant differences were found in any of the wavelengths. These data were confirmed by the analysis of the fit parameters. None of the fit parameters in the different datasets were affected by neoadjuvant chemotherapy, except for the fit parameter that reflects the amount of collagen, which showed a significant difference in only one dataset that comprised the measurements of tumor cells in combination with connective tissue. These findings seem to indicate that the impact of neoadjuvant chemotherapy on the optical characteristics of breast cancer tissue and healthy breast tissue types is limited, although some caution is warranted for drawing hard conclusions in the small numbered datasets. This means that also after neoadjuvant chemotherapy, margin assessment by DRS during breast-conserving surgery seems feasible.

### Disclosures

One of the authors is affiliated with Philips Research; B. H. is an employee of Philips Research; however, B. H. has no financial

interest in the subject matter, materials, and/or equipment. None of the other authors have a financial relationship with Philips or have other conflicts of interest to disclose.

### Acknowledgments

The authors would like to thank all surgeons and nurses from the Department of Surgery and all pathologists and pathologist assistants from the Department of Pathology for their assistance in collecting the specimens. We also thank the NKI-AVL Core Facility Molecular Pathology and Biobanking for supplying NKI-AVL biobank material.

### References

1. S. C. J. Bosma et al., "Very low local recurrence rates after breast-conserving therapy: analysis of 8485 patients treated over a 28-year period," *Breast Cancer Res. Treat.* **156**, 391–400 (2016).
2. M. G. Valero et al., "Surgeon variability and factors predicting for reoperation following breast-conserving surgery," *Ann. Surg. Oncol.* **25**, 2573–2578 (2018).
3. R. E. Pataky and C. R. Baliski, "Reoperation costs in attempted breast-conserving surgery: a decision analysis," *Curr. Oncol.* **23**(5), 314–321 (2016).
4. L. E. McCahill et al., "Variability in reexcision following breast conservation surgery," *J. Am. Med. Assoc.* **307**(5), 467–475 (2012).
5. A. J. Isaacs et al., "Association of breast conservation surgery for cancer with 90-day reoperation rates in New York state," *JAMA Surg.* **151**, 648–655 (2016).
6. L. G. Wilke et al., "Repeat surgery after breast conservation for the treatment of stage 0 to II breast carcinoma a report from the National Cancer Data Base, 2004–2010," *JAMA Surg.* **149**(12), 1296–1305 (2014).
7. D. J. Evers et al., "Diffuse reflectance spectroscopy: towards clinical application in breast cancer," *Breast Cancer Res. Treat.* **137**, 155–165 (2013).
8. L. L. De Boer et al., "Fat/water ratios measured with diffuse reflectance spectroscopy to detect breast tumor boundaries," *Breast Cancer Res. Treat.* **152**(3), 509–518 (2015).
9. L. L. De Boer et al., "Towards the use of diffuse reflectance spectroscopy for real-time in vivo detection of breast cancer during surgery," *J. Transl. Med.* **16**, 367 (2018).
10. Z. Volynskaya et al., "Diagnosing breast cancer using diffuse reflectance spectroscopy and intrinsic fluorescence spectroscopy," *J. Biomed. Opt.* **13**(2), 024012 (2008).
11. B. S. Nichols et al., "A quantitative diffuse reflectance imaging (QDRI) system for comprehensive surveillance of the morphological landscape in breast tumor margins," *PLoS One* **10**(16), e0127525 (2015).
12. Y.-H. Yu et al., "Prediction of neoadjuvant chemotherapy response using diffuse optical spectroscopy in breast cancer," *Clin. Transl. Oncol.* **20**(4), 524–533 (2017).
13. A. Leproux et al., "Performance assessment of diffuse optical spectroscopic imaging instruments in a 2-year multicenter breast cancer trial," *J. Biomed. Opt.* **22**(12), 121604 (2017).
14. W. T. Tran et al., "Multiparametric monitoring of chemotherapy treatment response in locally advanced breast cancer using quantitative ultrasound and diffuse optical spectroscopy," *Oncotarget* **7**(15), 19762–19780 (2016).
15. E. Provenzano et al., "Standardization of pathologic evaluation and reporting of postneoadjuvant specimens in clinical trials of breast cancer: recommendations from an international working group," *Mod. Pathol.* **28**, 1185–1201 (2015).
16. S. Kennedy et al., "The effects of hormonal and chemotherapy on tumoral and nonneoplastic breast tissue," *Hum. Pathol.* **21**, 192–198 (1990).
17. F. Aktepe, N. Kapucuoğlu, and I. Pak, "The effects of chemotherapy on breast cancer tissue in locally advanced breast cancer," *Histopathology* **29**, 63–67 (1996).
18. J.-H. Chen et al., "Decrease in breast density in the contralateral normal breast of patients receiving neoadjuvant chemotherapy: MR imaging evaluation," *Radiology* **255**(1), 44–52 (2010).
19. F. Fan, "Evaluation and reporting of breast cancer after neoadjuvant chemotherapy," *Open Pathol. J.* **3**, 58–63 (2009).
20. S. Sahoo and S. C. Lester, "Pathology of breast carcinomas after neoadjuvant chemotherapy," *Arch. Pathol. Lab. Med.* **133**, 633–642 (2009).
21. H. Cain et al., "Neoadjuvant therapy in early breast cancer: treatment considerations and common debates in practice," *Clin. Oncol.* **29**, 642–652 (2017).
22. J. A. Van der Hage, C. C. J. H. Van de Velde, and S. J. S. D. Mieog, "Preoperative chemotherapy for women with operable breast cancer (review)," *Cochrane Database Syst. Rev.* **2**, CD005002 (2007).
23. A. Charehbili et al., "Neoadjuvant hormonal therapy for endocrine sensitive breast cancer: a systematic review," *Cancer Treat. Rev.* **40**, 86–92 (2014).
24. A. E. Cerussi et al., "Diffuse optical spectroscopic imaging correlates with final pathological response in breast cancer neoadjuvant chemotherapy," *Philos. Trans. R. Soc. A* **369**, 4512–4530 (2011).
25. L. M. Arthur et al., "Pre-operative endocrine therapy," *Curr. Breast Cancer Rep.* **9**, 202–209 (2017).
26. H. Charfare, "Neoadjuvant chemotherapy in breast cancer," *Br. J. Surg.* **92**, 14–23 (2005).
27. L. L. De Boer et al., "Using DRS during breast conserving surgery: identifying robust optical parameters and influence of inter-patient variation," *Biomed. Opt. Express* **7**(12), 5188–5200 (2016).
28. R. Nachabé et al., "Estimation of lipid and water concentrations in scattering media with diffuse optical spectroscopy from 900 to 1600 nm," *J. Biomed. Opt.* **15**(3), 037015 (2010).
29. R. Nachabé et al., "Estimation of biological chromophores using diffuse optical spectroscopy: benefit of extending the UV-VIS wavelength range to include 1000 to 1600 nm," *Opt. Express* **1**(5), 1432–1442 (2010).
30. L. L. De Boer et al., "Method for co-registration of optical measurements of breast tissue with histopathology: the importance of accounting for tissue deformations," *J. Biomed. Opt.* **24**(7), 075002 (2019).
31. J. A. Hanley et al., "Statistical analysis of correlated data using generalized estimating equations: an orientation," *Am. J. Epidemiol.* **157**(4), 364–375 (2003).
32. J. Shults and S. J. Ratcliffe, "Analysis of multi-level correlated data in the framework of generalized estimating equations via xtmultcorr procedures in Stata and qls functions in MATLAB," *Stat. Interface* **2**, 187–196 (2009).
33. P. Taroni et al., "Breast tissue composition and its dependence on demographic risk factors for breast cancer: non-invasive assessment by time domain diffuse optical spectroscopy," *PLoS One* **10**(6), e0128941 (2015).
34. K. M. Blackmore et al., "The association between breast tissue and optical content and mammographic density in pre- and post-menopausal women," *PLoS One* **10**(1), e0115851 (2015).
35. E. Kho et al., "Hyperspectral imaging for resection margin assessment during surgery," *Clin. Cancer Res.* **25**(12), 3572–3580 (2019).
36. P. Taroni et al., "Non-invasive optical estimate of tissue composition to differentiate malignant from benign breast lesions: a pilot study," *Sci. Rep.* **7**, 40683 (2017).
37. R. Nachabé et al., "Diagnosis of breast cancer using diffuse optical spectroscopy from 500 to 1600 nm: comparison of classification methods," *J. Biomed. Opt.* **16**(8), 087010 (2011).
38. T. M. Bydlon et al., "Chromophore based analyses of steady-state diffuse reflectance spectroscopy: current status and perspectives for clinical adoption," *J. Biophotonics* **8**(1–2), 9–24 (2015).
39. O. Falou et al., "Diffuse optical spectroscopy evaluation of treatment response in women with locally advanced breast cancer receiving neoadjuvant chemotherapy," *Transl. Oncol.* **5**(4), 238–246 (2012).
40. T. D. O'Sullivan et al., "Optical imaging correlates with magnetic resonance imaging breast density and reveals composition changes during neoadjuvant chemotherapy," *Breast Cancer Res.* **15**, R14 (2013).
41. M. V. Pavlov et al., "Multimodal approach in assessment of the response of breast cancer to neoadjuvant chemotherapy," *J. Biomed. Opt.* **23**(9), 091410 (2018).
42. A. Sadeghi-Naini et al., "Early detection of chemotherapy-refractory patients by monitoring textural alterations in diffuse optical spectroscopic images," *Med. Phys.* **42**(11), 6130–6146 (2015).

43. A. M. Laughney et al., "Spectral discrimination of breast pathologies *in situ* using spatial frequency domain imaging," *Breast Cancer Res.* **15**, R61 (2013).
44. M. D. Keller et al., "Autofluorescence and diffuse reflectance spectroscopy and spectral imaging for breast surgical margin analysis," *Lasers Surg. Med.* **42**, 15–23 (2010).
45. J. A. Harvey et al., "Histologic changes in the breast with menopausal hormone therapy use: correlation with breast density, estrogen receptor, progesterone receptor, and proliferation indices," *Menopause* **15**(1), 67–73 (2008).
46. T. J. Anderson et al., "Effect of neoadjuvant treatment with anastrozole on tumour histology in postmenopausal women with large operable breast cancer," *Br. J. Cancer* **87**, 334–338 (2002).
47. W. R. Miller et al., "Pathological features of breast cancer response following neoadjuvant treatment with either letrozole or tamoxifen," *Eur. J. Cancer* **39**, 462–468 (2003).

**Lisanne L. de Boer** received her MSc degree in technical medicine from the University of Twente in 2013. Recently, she finished her PhD at the Netherlands Cancer Institute—Antoni van Leeuwenhoek Hospital. This project was focused on the use of fiberoptic DRS for detecting positive resection margins during surgery to improve the surgical treatment of breast cancer.

**Esther Kho** is a PhD candidate at the Netherlands Cancer Institute—Antoni van Leeuwenhoek Hospital. She received her MS degree in biomedical engineering from the University of Groningen in 2014. Her research involves the development, testing, and clinical evaluation of a hyperspectral imaging system, specifically for the assessment of tumor-positive resection margins during breast-conserving surgery.

**Katarzyna Józwiak** was working as a statistician at the Netherlands Cancer Institute—Antoni van Leeuwenhoek Hospital until June 2019. Currently, she is affiliated with the Institute of Biostatistics and Registry Research of the Brandenburg Medical School Theodore Fontane. In 2014, she obtained her PhD in applied statistics at Utrecht University.

**Koen K. Van de Vijver** is a surgical pathologist with an interest in diagnosis and treatment of cancer of the female genital tract and the breast. He received his PhD at Antwerp University and Leiden UMC in 2007. In 2008, he became an assistant professor at Maastricht University. After working as a consultant pathologist in the NKI-AVL,

he became a professor of gynaecological pathology at Ghent University Hospital in 2018.

**Marie-Jeanne T. F. D. Vrancken Peeters** has been working as a surgical oncologist specialized in breast cancer care in the Netherlands Cancer Institute—Antoni van Leeuwenhoek for more than 15 years now. She combines clinical activities with research activities with the main focus on development of localization techniques to facilitate breast cancer surgery, with the aim to de-escalate treatment whenever possible. Another area of interest is quality of breast cancer care, thus treating with the least amount of comorbidity.

**Frederieke van Duijnhoven** is a breast surgeon at the Netherlands Cancer Institute—Antoni van Leeuwenhoek Hospital. Her research is focused on de-escalating local treatment by adequate staging postneoadjuvant systemic therapy, minimizing resection volumes by accurate localization and by adopting a wait-and-see strategy in premalignant breast lesions.

**Benno H. W. Hendriks** is a researcher at Philips Research Eindhoven, The Netherlands, and professor in optics for minimally invasive instruments at the Department of Biomechanical Engineering, Delft University of Technology. He received his MSc degree in theoretical physics at RU Utrecht in 1985 and PhD in quantum optics at RU Utrecht in 1989. His current work focuses on tissue sensing for biomedical applications, image-guided therapy solutions, and minimally invasive devices for medical applications.

**Henricus J. C. M. Sterenborg** is active in biomedical optics research since 1987 as a staff member of the Medical Laser Centre of the Amsterdam UMC. In 1998, he co-founded the Centre for Optical Diagnostics and Therapy at the Erasmus Medical Center and in 2008 he became a professor of photodynamic therapy. Since 2013, he has held a joint position at the Department of Biomedical Engineering and Physics at the Amsterdam UMC and the Department of Surgery at the Netherlands Cancer Institute—Antoni van Leeuwenhoek Hospital.

**Theo J. M. Ruers** is an oncological surgeon and head of the Department of Surgery at the Netherlands Cancer Institute—Antoni van Leeuwenhoek Hospital. In 2010, he became a professor at the Faculty of Science and Technology at the University of Twente. He has extensive experience in oncologic surgery and the implementation of techniques in cancer care. Special focus is on using optical (imaging) techniques for diagnosis and treatment of cancer.